



Chloragard⁺ Antimicrobial/Antithrombogenic Catheter Technology Information

Introduction and Rationale for Antimicrobial Catheters:

Infection is the leading complication associated with intravascular devices, and there is a strong need to develop products to help prevent complications and increase safety for patients and providers. The National Nosocomial Infection Surveillance System (NNIS) tracks central line-associated bloodstream infection (BSI) rates in adult and pediatric intensive care units from 300 participating hospitals. This report serves as a benchmark for other hospitals to use in comparing their rates with the national rates. Approximately 90% of catheter-related bloodstream infections (CRBSIs) occur with central lines.⁸ Mortality attributable to CRBSIs has been reported to be between 4% to 20% resulting in prolonged hospitalization (a mean stay of 7 days) and increased hospital costs. Peripherally Inserted Central Catheters (PICCs) are associated with similar rates of CRBSI as Central Venous Catheters (CVCs), placed in internal jugular or subclavian veins (2 to 5 per 1,000 catheter days).¹¹

Vascular catheter infections develop for many reasons. They begin when a catheter becomes colonized by microorganisms entering through one or both of two routes: 1) colonization of the outside surface of the catheter or 2) colonization of the inside surface of the catheter. This colonization may be caused by any of five sources: environmental contamination, skin organisms, post-placement subcutaneous tract infection, intraluminal contamination or hematogenous seeding.¹²

Introduction and Rationale for Antithrombogenic Catheters:

Clinically symptomatic and detectable catheter-related venous thrombosis rates associated with peripherally inserted central venous catheters range from 3.4% (Turcotte, 2006) to as high as 20%.¹⁴ However, when diagnostic methods (ultrasound, contrast injection, etc) are used to assess for asymptomatic venous thrombosis, the incidence dramatically increases up to 58%.¹⁴ Occlusive and/or thrombotic events of peripherally inserted central venous catheters, described as inability to infuse solutions or withdraw blood, have an incidence of 7 to 25%.⁶

Catheter-related thrombus can be distinguished as either intraluminal, with clots occurring inside the lumen of the catheter, or extraluminal, with clots outside of the catheter and within the blood vessel (vein thrombosis). Formation of clot in the catheter lumen can lead to loss of its patency. If left untreated, extraluminal clot can cause complete occlusion of the blood vessel and can lead to a serious clinical condition called Deep Vein Thrombosis (DVT). The introduction of a venous catheter into the bloodstream triggers host responses to the presence of a foreign body. These host/biomaterial interactions occur on the external surface of the catheter, the internal surface of the venous wall, and the luminal surface of the catheter. The interactions of blood components, primarily proteins, platelets, and white blood cells in contact with the catheter material occur in a sequence of events. Within seconds of the catheter's exposure to the blood, protein adsorption and contact activation occur, followed by platelet adhesion, complement activation, and leukocyte adhesion minutes to hours later. The adhered bacteria, platelets and White Blood Cells (WBCs) become enmeshed within layers of fibrin forming a sheath on the surface of the catheter.

Technology Development:

Antimicrobial CVCs were introduced by Arrow International in 1990. The Arrow[®] catheter was the first commercially successful catheter capable of significantly reducing the potential for catheter colonization and subsequent catheter-related bloodstream infections.⁷ The first generation antimicrobial surface treatment, referred to as ARROWgard Blue[®], consists of two antimicrobial agents (chlorhexidine and silver sulfadiazine) which are impregnated into the indwelling external surface of the catheter. This combination has demonstrated broad spectrum *in vitro* efficacy as well as *in vivo* efficacy through prospective clinical studies.^{1,4,5,8,17}

Due to the need for longer duration of protection as a result of longer dwelling time and in recognition of the role of the intraluminal pathway in catheter colonization by organisms transmitted by the hands of unit personnel,^{7,12,13} two key areas of improvement to the ARROWgard Blue catheter technology were identified: 1) extend the effective duration of action of the external surface coating and 2) provide protection to the internal surfaces of the entire

catheter (including extension lines and hubs). The second generation antimicrobial catheter, known as ARROWg[†]ard Blue PLUS[®] (AGB^{†*}), was developed to address these needs. This was done by increasing chlorhexidine on the outside surface of the catheter and also by protecting the entire intraluminal path with chlorhexidine. Compared to the original ARROWg[†]ard Blue, ARROWg[†]ard Blue PLUS catheters produce a significantly longer duration of antimicrobial effect against the most common catheter-related infection-causing microorganisms, including a significant reduction in intraluminal bacterial colonization when compared to untreated catheters.⁹

The third generation of antimicrobial catheter technology was introduced on catheters as Chlorag[†]ard[®] Technology, with slight modification to the clinically proven efficacy of the ARROWg[†]ard Blue PLUS technology. Silver sulfadiazine, the secondary antimicrobial agent, has been removed, and chlorhexidine-to-catheter material processing was optimized to provide longer duration based on the clinical requirements of PICCs and Jugular Axillo-subclavian Central Catheters (JACC[™]). This technology has been shown to have antithrombogenic properties as well.

Product Description:

Catheters with Chlorag[†]ard[®] Technology are processed with an external surface treatment that uses antimicrobial chlorhexidine acetate on the catheter body and juncture hub nose, as well as an internal lumen impregnation utilizing an antimicrobial combination of chlorhexidine acetate and chlorhexidine base for the catheter body, juncture hub, extension line(s) and extension line hub(s). The maximum total amount of chlorhexidine applied to a 55 cm length catheter may be up to 18.6 mg. Other lengths of PICCs and all lengths of JACCs are less than 55 cm, and will contain less chlorhexidine than the 55 cm length.

Characterization of Chlorhexidine:

Chlorhexidine is characterized as having a broad antimicrobial activity spectrum, including bacteriostatic and bactericidal effects on gram positive bacteria, gram negative bacteria and fungi. Chlorhexidine also has been shown to be effective against viruses with a lipid component in their coats or with an outer envelope, but these properties have not been evaluated with this product.

The antithrombogenic effect of Chlorag[†]ard[®] Technology on catheters appears to be a function of thrombin inhibition by chlorhexidine via intrinsic and common pathways of blood coagulation, causing delayed blood clotting response and thrombus accumulation on catheter surface.

Whether chlorhexidine is bacteriostatic or bactericidal depends largely on the concentration of the agent, its pH and the susceptibility of specific organisms.^{2,18}

Chlorhexidine is a cationic compound. Its positively charged molecules are strongly attracted to the negative surface charges of bacterial cells. The outer membrane of gram negative bacteria, cell wall of gram positive bacteria or cytoplasmic membrane of yeasts then becomes weakened from increased permeability caused by chlorhexidine being adsorbed onto the cell surface. Chlorhexidine exhibits bacteriostatic effects at low concentrations due to the release of substances characterized by low molecular weights (i.e., phosphorus and potassium ions) from the cell. This damage is enough to inhibit bacterial cell function. Bactericidal activity of chlorhexidine occurs at higher concentrations by causing precipitation of proteins and nucleic acids.²

Chlorhexidine is poorly absorbed from the gastrointestinal tract. In human and animal studies, the average plasma level peaked at 0.206 µg in humans 30 minutes after ingesting 300 mg of chlorhexidine. Excretion occurred primarily through the feces (about 90%), and less than 1% was excreted in urine. Chlorhexidine is metabolized in the same manner as most other foreign substances. The majority will be excreted without being metabolized.²

Preclinical biocompatibility studies support the conclusion that there is a negligible risk of adverse effects from Chlorag[†]ard[®] antimicrobial/antithrombogenic catheters.

Indications for Use:

Chlorag[†]ard[®] Technology treatment on the external surface of the catheter body as well as the entire fluid pathway of the catheter has been shown to be effective in reducing microbial colonization on catheter surfaces. Antimicrobial effectiveness was evaluated using in vitro and in vivo test methods and no correlation between these testing methods and clinical outcome has currently been ascertained. It is not intended to be used for the treatment of existing infections.

Contraindications:

The Chloragard antimicrobial / antithrombogenic catheter is contraindicated for patients with known hypersensitivity to chlorhexidine.

Warning:

Remove catheter immediately if adverse reactions occur after catheter placement.

NOTE: Perform sensitivity testing to confirm allergy to catheter antimicrobial agents if adverse reaction occurs.

Refer to enclosed product Instructions for Use (IFU) for additional Warnings and Precautions.

Hypersensitivity Potential:

Benefits of the use of this catheter should be weighed against any possible risk. Hypersensitivity reactions are a concern with antimicrobial catheters and can be serious and even life-threatening. Since antimicrobial catheters were introduced into the market, there have been some reports of hypersensitivity occurrences outside the United States. This hypersensitivity potential has been reported to occur more frequently in Japan.

Pre-Clinical Evaluations:

Chloragard Technology has demonstrated microbial colonization reduction against gram-positive, gram-negative and yeast in *in vitro* and *in vivo* studies for up to 30 days for external surface and *in vitro* studies for up to 30 days for fluid pathway.¹⁶

In addition, Chloragard Technology has also demonstrated reduction in thrombus accumulation on catheter surfaces for up to 30 days in *in vivo* testing. *In vitro* testing has exhibited reduction in platelet adhesion on catheter surface and catheter occlusion.¹⁶

Clinical Evaluations:

Reduction in colonization or microbial growth on Chloragard catheters has not been shown to correlate with a reduction in infections in patients. Clinical studies to evaluate reduction in infection have not been performed on these devices. Clinical effectiveness of Chloragard catheters in preventing CRBSIs compared to ARROWgard Blue PLUS catheters has not been studied. The coating on both products primarily contains the antimicrobial agent chlorhexidine with similar concentration per surface area, which has been shown to be effective in reduction of colonization of catheter surfaces in *in vitro* testing. Based on similarities of the Chloragard antimicrobial / antithrombogenic catheters with ARROWgard Blue PLUS catheter technology and clinical usage, the studies performed

on ARROWgard Blue PLUS antimicrobial catheters listed below may provide a useful comparison in demonstrating clinical safety and effectiveness of the Chloragard Technology in patients.

Clinical Study - France³

A prospective, multi-center, randomized, double-blind clinical study of 397 patients performed at 14 university-affiliated hospital ICUs in France from June 1998 to January 2002 using ARROWgard Blue PLUS antimicrobial catheters showed use of these catheters was associated with a strong trend toward reduction in infection rates of central venous catheters (colonization rate of 3.7% versus 13.1%, 3.6 versus 11 per 1000 catheter-days, $p=0.01$) and CVC-related infection (bloodstream infection) in 4 versus 11 (2 versus 5.2 per 1000 catheter-days, $p=0.10$).

Clinical Study - Germany⁹

A prospective, randomized, double-blind, controlled clinical study of 184 patients performed at the University Hospital of Heidelberg (Heidelberg, Germany) from January 2000 to September 2001 using ARROWgard Blue PLUS antimicrobial catheters showed these catheters were effective in reducing the rate of significant bacterial growth on either the tip or subcutaneous segment (26%) compared to control catheters (49%). Incidence of catheter colonization was also significantly reduced (12% coated versus 33% uncoated). The number of bloodstream episodes in patients with CHSS catheter was lower than in patients provided with control catheter (3 versus 7 episodes, $p=0.21$).

Clinical Study - United States¹⁰

A prospective, multi-center, randomized, double-blind, controlled clinical study of 780 patients performed at 9 university-affiliated hospitals in the United States from July 1998 to June 2001 using ARROWgard Blue PLUS antimicrobial catheters showed these catheters were less likely to be colonized at time of removal compared to control catheters (13.3 versus 24.1 colonized catheters per 1000 catheter-days, $p<0.01$). Rate of definitive catheter-related bloodstream infection was 1.24 per 1000 catheter days (CI, 0.26 to 3.26 per 1000 catheter-days) for control group versus 0.42 per 1000 catheter days (CI, 0.01 to 2.34 per 1000 catheter-days) for ARROWgard Blue PLUS catheter group ($p=0.06$).

No adverse events were observed from ARROWgard Blue PLUS catheters in any of the clinical studies.

Refer to enclosed product Instructions for Use (IFU) for specific indications, procedural technique(s) and potential complications associated with PICC or JACC insertion procedures.

References:

1. Bach A, Bohrer H, Bottiger B, Motsch J, Martin E. Reduction of bacterial colonization of triple-lumen catheters with antiseptic bonding in septic patients. *Anesthesiology*. 1994;81:A261.
2. Block, S.S. 2001. Chlorhexidine. Ch. 15 in *Disinfection, Sterilization and Preservation*, 5th ed., S.S. Block (Ed.), p. 321-336, 2001.
3. Brun-Buisson C, Doyon F, Sollet J, Cochar J, et al. Prevention of intravascular catheter-related infection with newer chlorhexidine-silver sulfadiazine-coated catheters: a randomized controlled trial. *Intensive Care Medicine*. 2004;30:837-843.
4. Civetta J, Hudson-Civetta J, Ball S. Decreasing catheter related infection and hospital costs by continuous quality improvement. *Crit Care Med*. 1996;24:1660-5.
5. Collin GR, Decreasing catheter colonization through the use of an antiseptic-impregnated catheter: a continuous quality improvement project. *Chest*. 1999;115:1632-1640.
6. Hoffer, E. K., Borsa, J., Santulli, P., Bloch, R., & Fontaine, A. B. (1999). Prospective randomized comparison of valved versus nonvalved peripherally inserted central vein catheters. *American Journal of Roentgenology*, 173(5), 1393-1398
7. Maki DG, Ringer M. Evaluation of dressing regimes for prevention of infection with peripheral intravenous catheters. *JAMA*. 1987;258:2396-2403.
8. Maki DG, Stolz SM, Wheeler S, Mermel LA. Prevention of central venous catheter-related bloodstream infection by use of an antiseptic-impregnated catheter. *Ann Intern Med*. 1997;127:257-266.
9. Ostendorf T, Meinhold A, Harter C, Salvwender H, et al. Chlorhexidine and silver sulfadiazine coated central venous catheters in haematological patients – a double-blind, randomized, prospective, controlled trial. *Support Care Cancer*. 2005;13:993-1000.
10. Rupp M, Lisco S, Lipsett P, Perl T, et al. Effect of a second-generation venous catheter impregnated with chlorhexidine and silver sulfadiazine on central catheter-related infections. *Annals of Internal Medicine*. October 18, 2005;143(8):570-581.
11. Safdar, N and Make DG. Risk of Catheter-Related Bloodstream Infection With Peripherally Inserted Central Venous Catheters Used in Hospitalized Patients. *Chest*. 2005; 285: 489-495.
12. Sherertz RJ. Pathogenesis of vascular catheter-related infections. In: Seifert H, Jansen B, Farr BM, eds. *Catheter-Related Infections*. New York, NY: Marcel Dekker, Inc; 1997:1-29.
13. Sherertz RJ, Heard SO, Raad II. Diagnosis of triple-lumen catheter infection: comparison of roll plate, sonication, and flushing methodologies. *J Clin Microbiol*. 1997;35:641-646
14. Trerotola, S, Stavropoulos, S, Mondschein, J, et al. Triple-lumen peripherally inserted central catheter in patients in the critical care unit: prospective evaluation. *Radiology*. 2010; 256(1): 312-330
15. Turcotte, S, Dube', S, Beauchamp, G. Peripherally inserted central venous catheters are not superior to CVCs in the acute care of surgical patients on the ward. *World Journal of Surgery*. 2006; 30:1605-1619
16. Testing Performed by Independent Laboratories: data on file at Arrow International.
17. Veenstra DL, Saint S, Somnath S, Lumley T, Sullivan SD. Efficacy of Antiseptic-Impregnated Central Venous Catheters in Preventing Catheter-Related Bloodstream Infection. *JAMA*. 1999;281:261-267.
18. Xu QA, Zhang, Y, Trissel, LA, et al. Adequacy of a New Chlorhexidine-Bearing Polyurethane Central Venous Catheter for Administration of 82 Selected Parenteral Drugs. *Annals of Pharmacotherapy*. October 2000; 13: 1109-1116.

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